un evento promosso da

ISTITUT ROMAGN LO PER L STUDIO DEI TUMORI DINO AMADOR

Nuove frontiere del Next Generation Sequencing nella diagnostica oncologica ed ematologica 04 Novembre 2022 Centro Congressi FEDERICO II Napoli

ANTONIO PINTO - STEFANIA CRISCI

UOSC EMATOLOGIA ONCOLOGICA – INT NAPOLI– IRCCS – G. Pascale

L'NGS nei linfomi: il laboratorio incontra il clinico



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NGS IN LYMPHOMAS MAY ENABLE

- <u>diagnostic refinement</u> through multiple genomic biomarkers identified simultaneously (potentially challenging differential diagnoses)
- <u>risk stratification</u> (prognosis and therapy prediction)

... AND DISCLOSE

- <u>genomic alterations</u> (therapeutically targetable alterations and vulnerabilities)
- biomarkers of <u>drug resistance</u> and <u>real-time monitoring</u>, with early detection of relapse (opening the way for personalized medicine)
- a mutation database (a source for new drugs in lymphoma)



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DETECTION CAPACITY OF GENOMIC ABERRATIONS WITH DIFFERENT TECHNOLOGIES

		Single Nucleotide Variants/ InDels	Copy Number Alterations ³	Structural Variants⁴	IG/TR Clonality	Cell of Origin	Tumor. Purity
_	Fluorescence <i>in situ</i> Hybridization		~	~			
etec	Single gene analyses ¹	~			~		
Targeted	Amplicon-based gene panel sequencing	~			~		
	Capture-based gene panel sequencing	~	∇	~	~		
ב מ	Genomic arrays		~				~
Digital/ Arrays	Methylation arrays		~			× .	~
	Gene expression ²					~	
e	Whole transcriptome sequencing	$\mathbf{\nabla}$			~	 Image: A start of the start of	
Genome Wide	Whole exome sequencing	~	$\mathbf{\nabla}$		~		~
e e	Whole genome sequencing	\checkmark	\checkmark	× .	\checkmark		~



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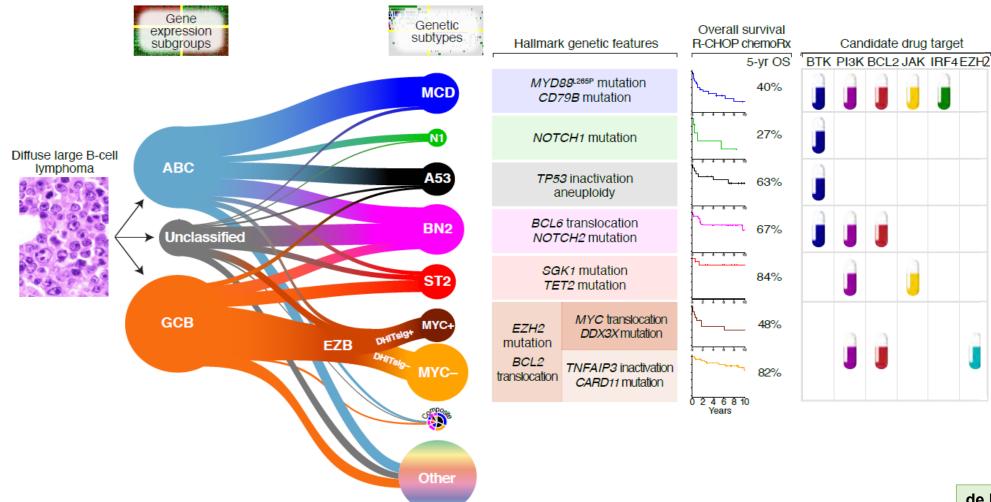
DNA METHYLATION AND CHROMATIN PROFILING

Epigenetic mechanisms play critical roles in lymphomagenesis and have significant clinical diagnostic and outcome implications. A DNA methylation imprint of the cellular origin is useful for diagnostic and patient stratification purposes.

- 1. Aberrant <u>histone modifications</u> are critically relevant to lymphomagenesis. Extensive changes in the activity of regulatory elements are targets of drugs such as BET inhibitors. For example
 - I. EZH2, after gain-of-function mutations, causes profound spreading of the H3K27me3 promoter repressive mark, which is reversed by EZH2 inhibitors
 - II. KMT2D loss-of-function mutations cause loss of enhancer-activating H3K4me1 and may be reverted through inhibition of histone demethylases
- 2. Recurrent <u>hypermethylation</u> of specific genes is harboured by Lymphoid neoplasms, including
 - . CDKN2A, a canonical tumour suppressor gene, is related to disease progression
 - **II. SMAD1** is a biomarker for chemotherapy resistance
 - III. TET2 may present epigenetic modifier mutations due to a hypermethylation effect that drives an aberrant cytosine methylation patterning, a universal finding in lymphoid neoplasm.

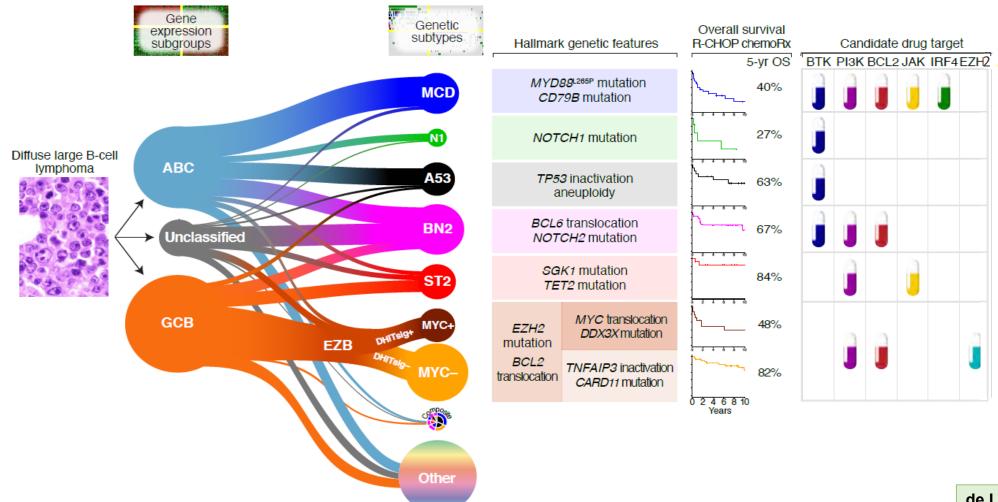
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APPROACH TO DIAGNOSING HIGH-GRADE B-CELL LYMPHOMAS

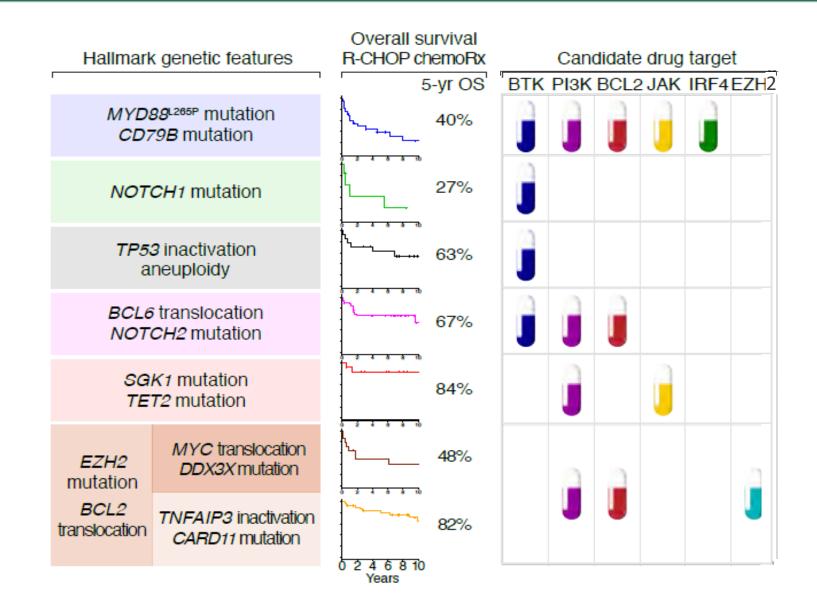


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APPROACH TO DIAGNOSING HIGH-GRADE B-CELL LYMPHOMAS



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NGS in Lymphoma Diagnosis

PRACTICAL CONSTRAINTS:

- 1. The optimal source is nucleic acids extracted from fresh surgical biopsies or liquid samples.
 - ⇒ Yet, clinical assays still need to be optimally fit for formalin-fixed paraffin-embedded (FFPE) tissues, the common diagnostic material. Also, fresh needle biopsies are suboptimal samples.
- 2. Large and complex panels by capture-based NGS are required to analyze chromosomal aberrations, CNA, SNV and INDEL simultaneously and to provide uniform coverage for sensitive detection of subclonal somatic abnormalities.

 \Rightarrow Still, amplicon target enrichment is applied as well.

3. Tumour cell content, gene selection, sequencing platform, sequence coverage/depth, background artefacts, unique molecular identifiers, variant interpretation and turnaround time are critical parameters for NGS-based assays.

 \Rightarrow Still, the procedures and the quality of data differ among laboratories.

4. A comparison of sequential biopsies may be necessary depending on the clinical question. ⇒ Still, only the most recent sample is available for analysis in case of disease recurrence.



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NGS in Lymphomas - Next steps

Validation & Standardization

There has yet to be a standard approach. Features such as gene selection, sequencing platform, read depth, and variant analysis can differ among laboratories

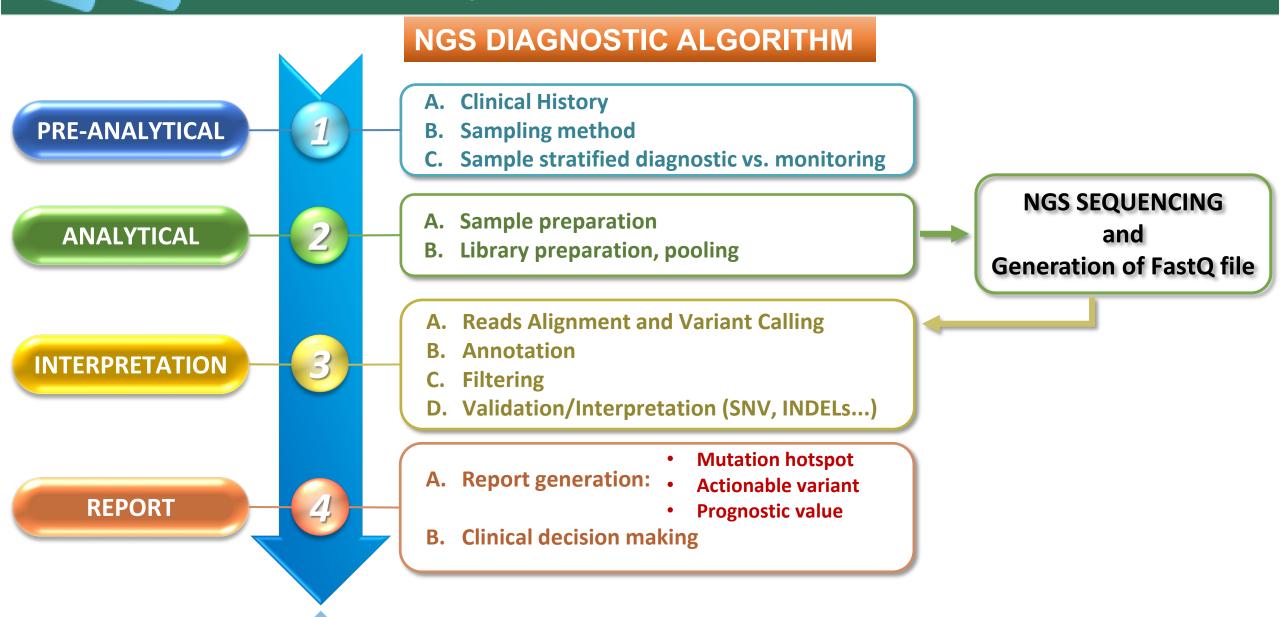
SOP

A standard operating procedure (SOP) for the classification of the oncogenicity of somatic variants should be devised

Liquid biopsies

The use of NGS on liquid biopsies will be a breakthrough not only towards tailormade therapies at diagnosis but also towards a real-time and dynamic monitoring of tumour responses to treatment

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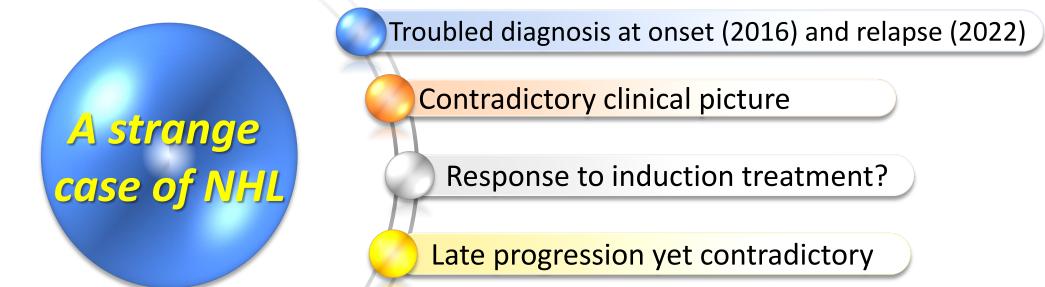
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Clinical Case

A STRANGE KIND OF NHL *«...between the stables and the 'starry sky'!»*

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Topics covered



Contribution of NGS and new therapy



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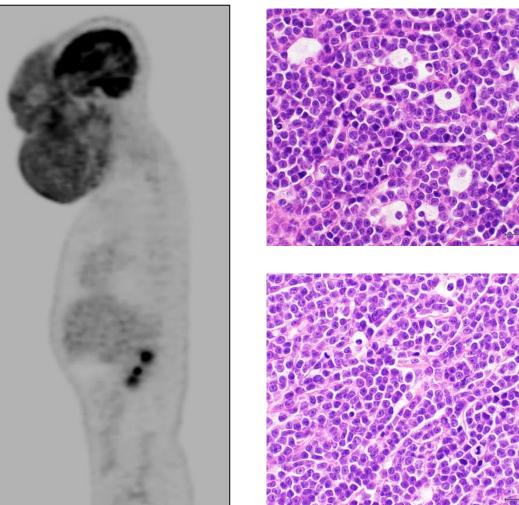
OCTOBER 2016, Multiple biopsies: Indolent NHL (Ivory Coast-France) vs. high suspicious for Burkitt lymphoma with t(8;14) and 'STARRY SKY' (Naples)



• Age: 17, Male • African ethnic origin



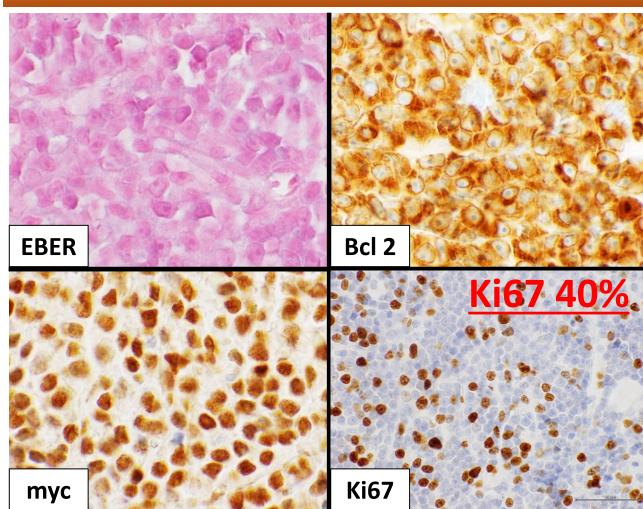


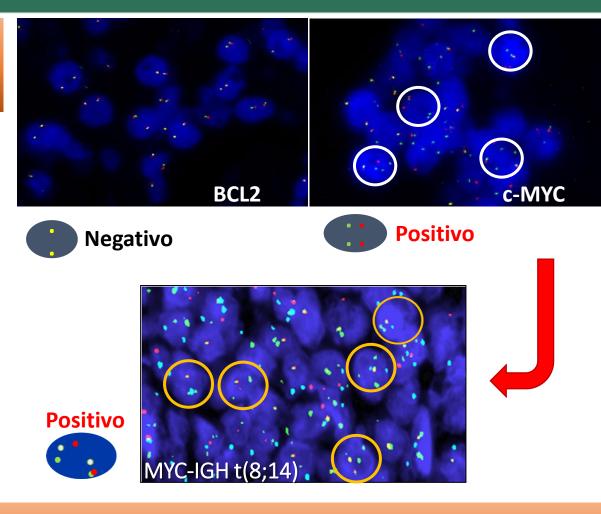




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IHC: CD20+CD10-,BCL6-, BCL2+(100%), MUM1+,c-myc+(100%),EBER-,Tdt-,Ki67 40%





FISH: MYC-IGH t(8;14): POSITIVE

PCR: IGH/BCL2 & IGH/CCND1 & API2/MALT1: all NEG., IGH rearrangement monoclonal



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Induction Treatment

R-CODOX-M/R-IVAC x 4 cycles, w/ SNC prophylaxes \rightarrow Partial response (only!)

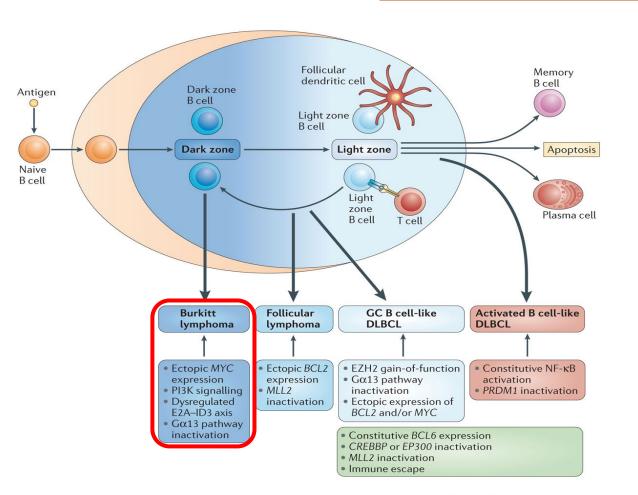
JAN 2017: 2nd line treatment w/ GDP (2 cycles) + IFRT followed by ASCT consolidation



Follow-up

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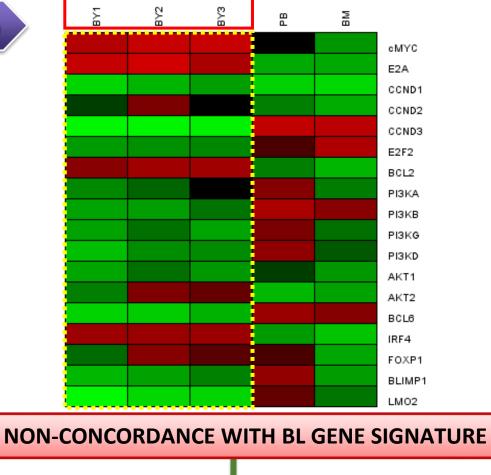
GENE EXPRESSION PROFILING





2 Step

-3.0



Sanger sequencing of c-MYC gene

1:1

3.0

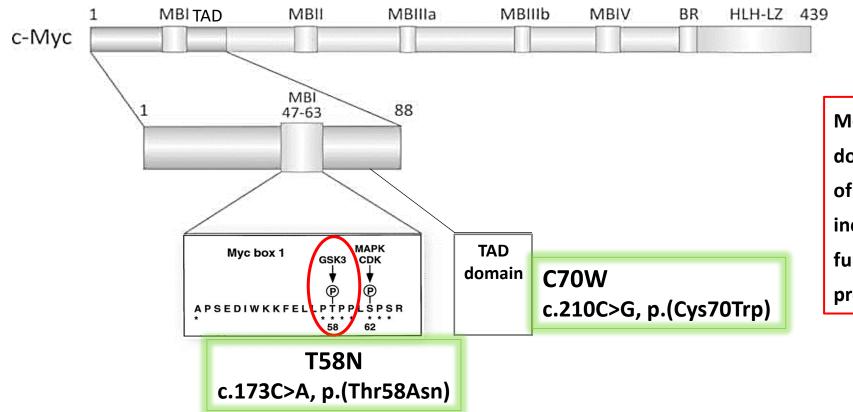
Nature Reviews | Immunology



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SEQUENCING OF c-MYC ONCOGENE

Paired-end Sanger sequencing with 34 primer pairs



Most of c-MYC mutations target functional domains that enhance the oncogenic potential of MYC by different mechanisms, including increased protein stability and transcriptional function, or by impairing the induction of the proapoptotic element BIM

c-MYC: NM_002467.4



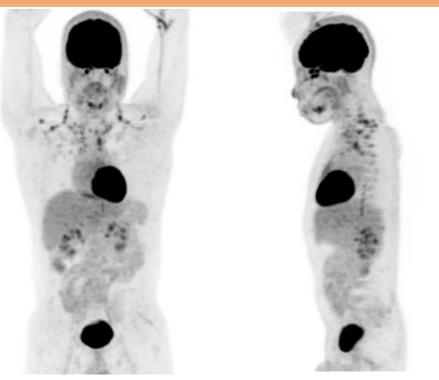
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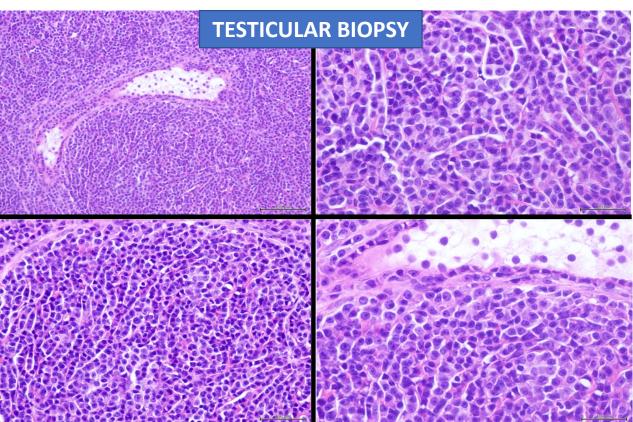
<u>Recurrence: September 2022</u> – Histology: «comparable to 2016 … HGL-NOS/WHO 2017, <u>Ki67→30 to 40% (transformation from a low-grade NHL?)</u>.»

Site of Recurrence:

PET NEGATIVE: mandibular, paravertebral (T5-T6, at RMN), testicular (PET-, ECO+, Hystology)

> PET POSITIVE: orbital (SUV 3.2)(Histology)



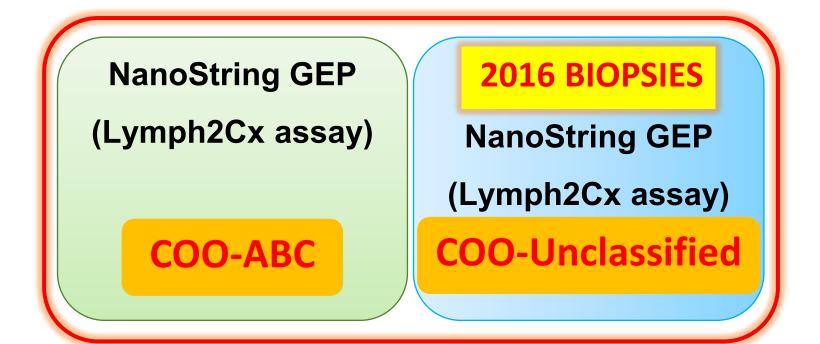




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2022 MOLECULAR PROFILING

PCR: IGH/BCL2 & IGH/CCND1 & API2/MALT1: ALL NEGATIVE, IgH GENE REARRANGEMENT: MONOCLONAL



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TARGETED SEQUENCING: CAPTURE-BASED NGS-LYMPHOMA SOLUTION PANEL

2022

Gene Transcript, CDNA, Protein	Type Consequence	Pathogenicity Classification	Orbital biopsy VF	Testicular biopsy VF	BMA VF	PB VF
CHD2 NM_001271, c.2636C>T, p.(Ala879Val)	SNP missense	Likely Pathogenic	44,8%	-	-	-
CHD2 NM_001042572, c.725C>A, p.(Ser242*)	SNP nonsense	VUS	-	47%	-	-
CHD2 NM_001271, c.4173dupA, p.(Gln1392Thrfs*17)	INDEL frameshift	VUS	5,1%	5,3%	-	-
KRAS NM_004985, c.37G>T, p.(Gly13Cys)	SNP missense	Pathogenic	48,8%	47,8%	-	-
MYC NM_002467, c.255C>G, p.(Cys85Trp)	SNP missense	Pathogenic	48%	51,6%	-	-
MYC NM_002467, c.218C>A, p.(Thr73Asn)	SNP missense	Pathogenic	47,8%	51,2%	-	-
IGH clonality-NGS			IGHV4-34*02- D3-10*01-J5*02	IGHV4-34*02- D3-10*01-J5*02	-	-



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TARGETED SEQUENCING: CAPTURE-BASED NGS-LYMPHOMA SOLUTION PANEL

2016							
Gene Transcript, CDNA, Protein	Type Consequence	Pathogenicity Classification	Perimandibular biopsy VF	Cheek biopsy VF	BMA VF	PB VF	
CHD2 NM_001271, c.4173dupA, p.(Gln1392Thrfs*17)	1271, c.4173dupA, frameshift VUS -		-	5,2%	-	-	
KRAS NM_004985, c.37G>T, p.(Gly13Cys)	SNP missense	Pathogenic	20,1%	20,9%	-	-	
MYC NM_002467, c.255C>G, p.(Cys85Trp)	SNP missense	Pathogenic	40,4%	38,3%	-	-	
MYC NM_002467, c.218C>A, p.(Thr73Asn)	SNP missense	Pathogenic	39,8%	38,7%	-	-	
IGH clonality-NGS			IGHV4-34*02/ D3-10*01/J5*02	IGHV4-34*02/ D3-10*01-J5*02	-	-	

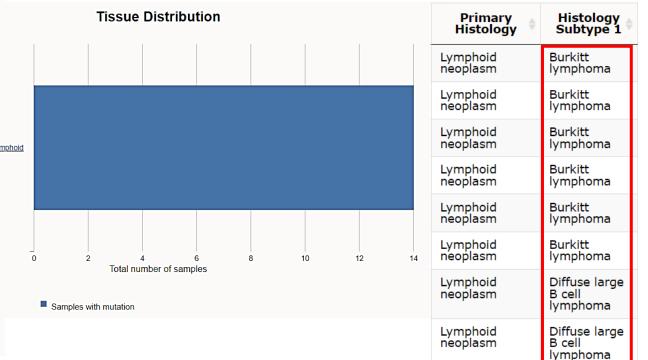
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SCOSMIC GRCh37

c-MYC Variant c.218C>A (p.T73N)

This variant is in protein domain (exon 2):
Transcription regulator Myc, N-terminal SIFT (v6.2.0): DELETERIOUS (1.0)

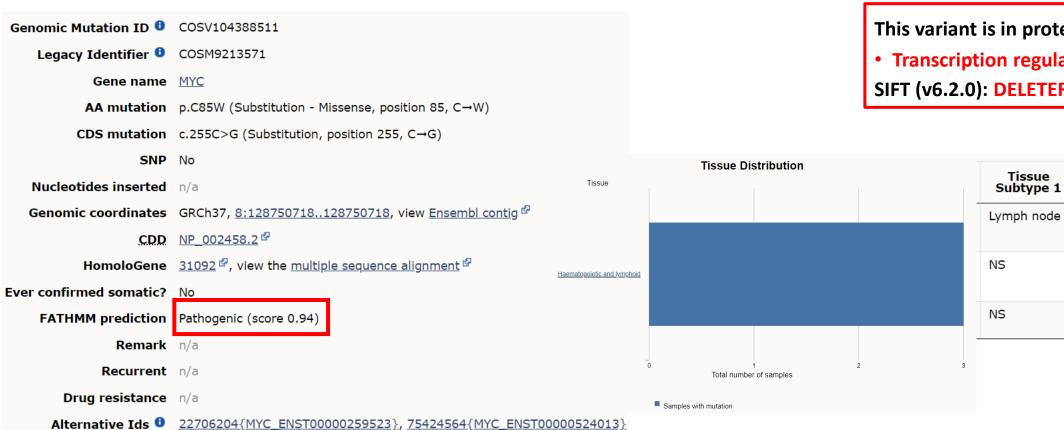




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SCOSMIC GRCh37

c-MYC Variant c.255C>G(p.C85W)



This variant is in protein domain (exon 2):

 Transcription regulator Myc, N-terminal SIFT (v6.2.0): **DELETERIOUS (0.99)**

Primary Histology

Lymphoid

neoplasm

Lymphoid

neoplasm

Lymphoid

neoplasm

Histology Subtype 1

Diffuse large

Diffuse large

lymphoma

lymphoma

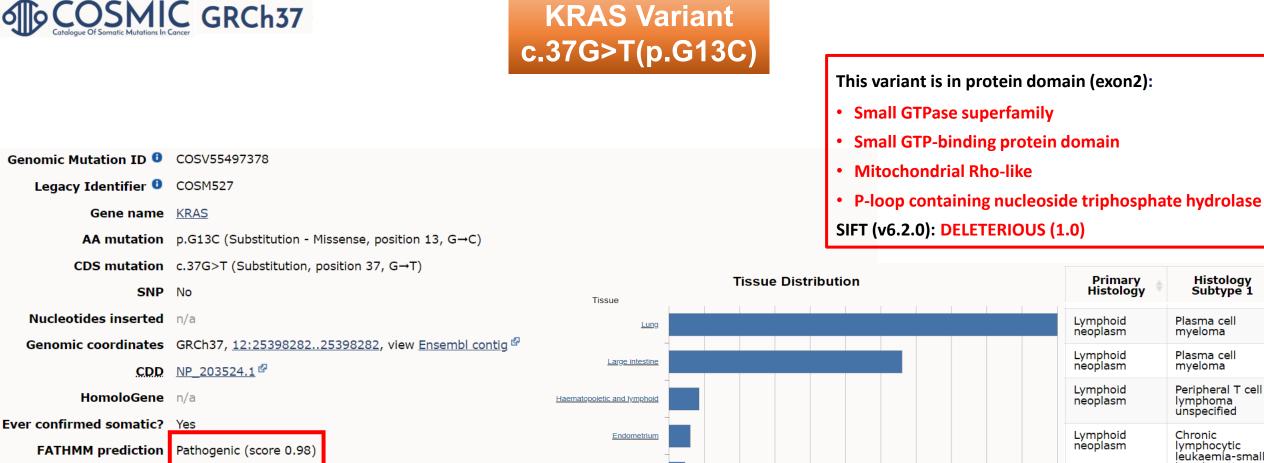
lymphoma

B cell

B cell

Burkitt

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Ovary

0

20

60

80

Total number of samples

40

100

120

140

160

180

200

220

Lymphoid Plasma cell néoplasm myeloma Lymphoid Plasma cell néoplasm myeloma Lymphoid Peripheral T cell lymphoma neoplasm

Primary

Histology

Lymphoid

néoplasm

neoplasm

neoplasm

Haematopoietic

Haematopoietic

Histology

Subtype 1

únspecified

lymphocytic

lýmphoma

leúkaemia

leukaemia

Chronic

leukaemia-small lymphocytic

myelomonocytic

Acute myeloid

Chronic

Samples with mutation Alternative Ids 0 29423429{KRAS ENST00000311936}, 88011751{KRAS ENST00000556131}, 87542567{KRAS ENST00000557334}

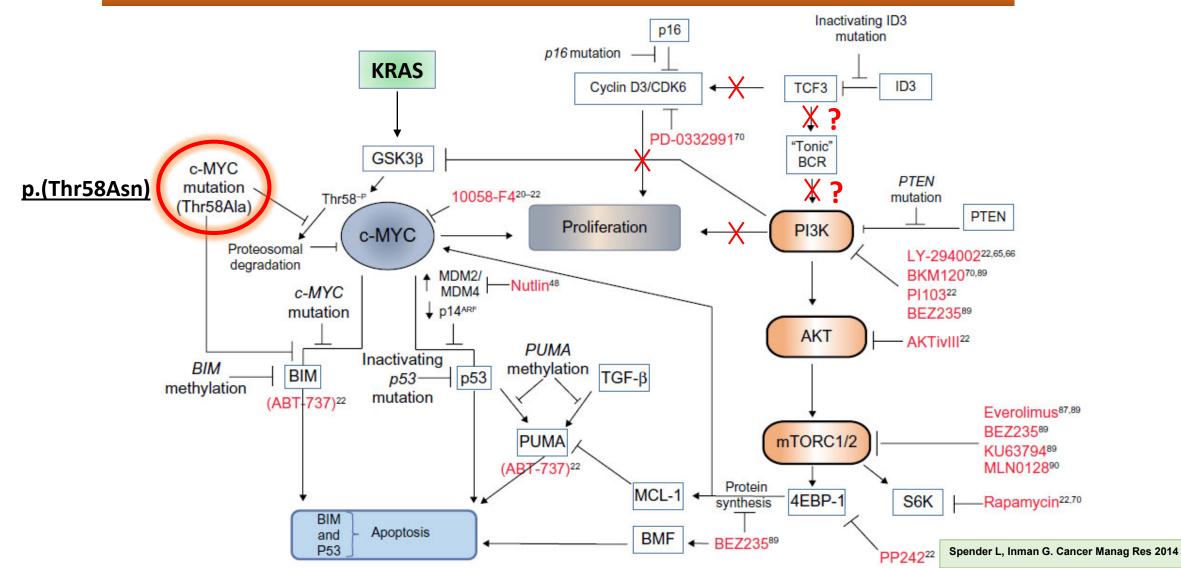
Remark n/a

Recurrent n/a

Drug resistance n/a

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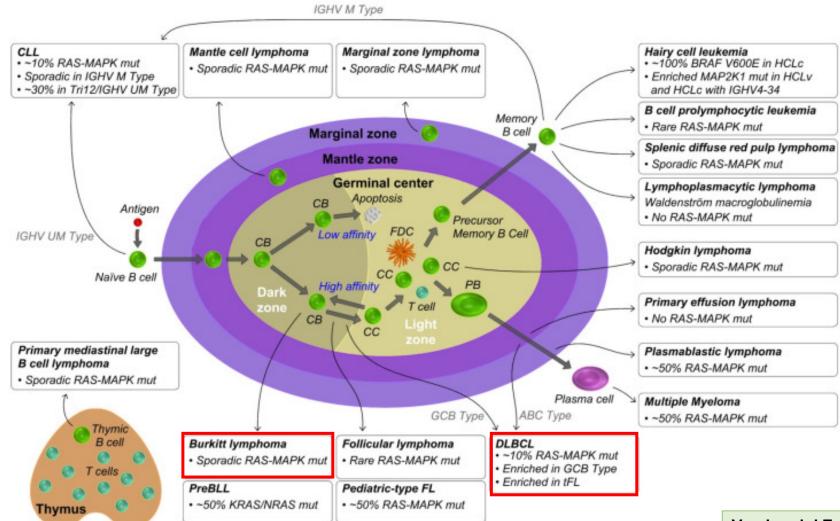
COOPERATING MUTATIONAL EVENTS IN MYC-DRIVEN LYMPHOMAGENESIS





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MATURE B CELL LYMPHOPROLIFERATIVE DISORDERS AND RAS-MAPK PATHWAY DEREGULATION

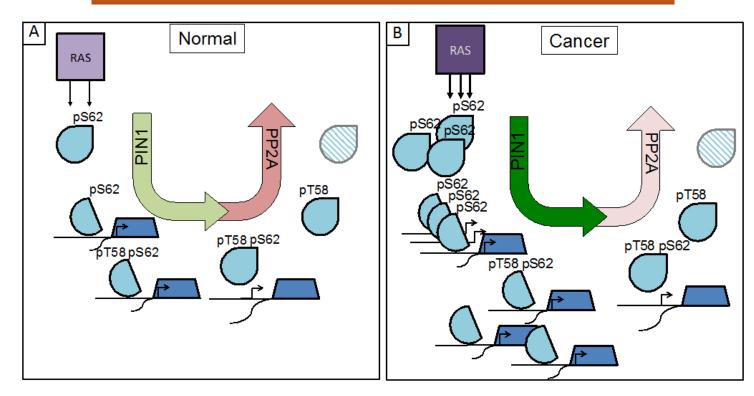


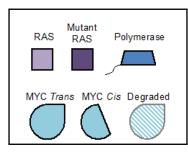
Vendramini E, et al. Cancers (Basel) 2022



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RAS AND MYC: CO-CONSPIRATORS IN CANCER





A. In normal cells, RAS signaling leads to MYC phosphorylation at S62, which supports PIN1 isomerization of P63 to cis, recruitment of MYC to target gene promoters, and activation of the basal transcription machinery. Subsequent phosphorylation of MYC at T58 results in a second PIN1 isomerization of P63 to trans that is associated with the release of MYC from DNA, PP2A mediated pS62 dephosphorylation and MYC degradation.

B. In RAS-driven cancer, increased signaling from mutant RAS along with active PIN1 and suppressed PP2A leads to an accumulation of active MYC that can drive pro-tumor transcriptional programs.



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LNH- MUTATIONAL SPECTRUM	CLINICAL CASE (MYC, KRAS, CHD2)			
T-HGL of indolent B-cell LNH (TNFAIP3, GPR34, CD274, TNFRSF14, TET2, EZH2, NOTCH2, NOTCH1, CREBBP, KMT2D, MYC, TP53)	\checkmark			
DLBCL NOS GCB (EZH2, GNA13, MEF2B, KMT2D, B2M, TNFRSF14, CREBBP)	\checkmark			
DLBCL NOS ABC (MYD88 p.L265P, CD79B, PIM1, PRDM1/BLIMP1, MYC)	\checkmark			
HGL NOS (MYD88, CD79B, TBL1XR1, MYC, KMT2D, TP53, BCL2, EZH2 CREBBP, TNFRSF14)	\checkmark			
BL SPORADIC (MYC, TCF3, ID3)	\checkmark			

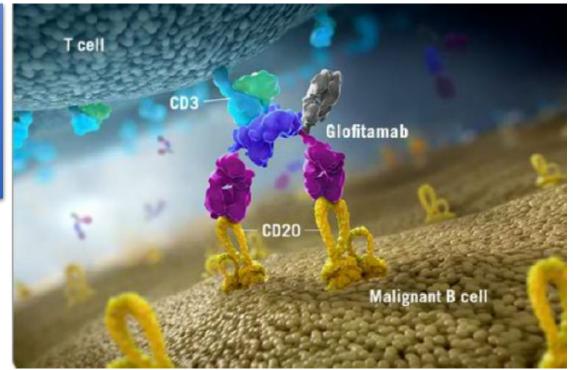


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«... the aggressive subtype documented through NGS prompted us to enrol the patient in an appropriate clinical trial...»

Phase 1 study NP39488 Polatuzumab (ADC, anti-CD79b) + Glofitamab (Bispecific Antibody, antiCD20xCD3)

NCT03533283 Espansion Part for relapsed/refractory **DLBCL and High-grade B-cell lymphoma**



glofitamab

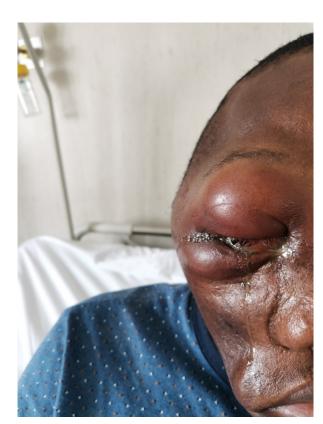
(CD20)2 x CD3

- humanized mouse IgG1-based antibody
- bivalent CD20 and monovalent CD3e binding
- modified Fc devoid of FcyR and complement binding

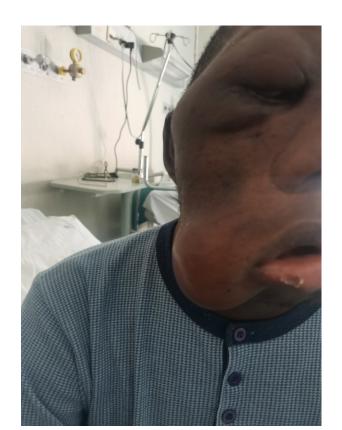


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Flares occurring 36 hours after 1° step-up dose of Glofitamab







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Clinical case – The contribution of NGS

Diagnosis

Despite renewed efforts on the histological classification of high-grade lymphomas, still exists a grey zone with clinical and pathological conundrums

Prognosis

Clinico-pathological prognostication tools do not adequately encompass the heterogeneities of high-grade lymphomas, while histological classification is far from remedying.

Therapy

NGS may retain diagnostic and prognostic value in high-grade NHL. Moreover, genomic alterations detectable through NGS may represent an additional source for therapeutically targetable alterations and vulnerabilities, well beyond immunotherapies such as glofitamab and polatuzumab adopted in this clinical case.

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Thankful Tree

